General comments for all tests: (from Helpful Hints 19 July 2004 handout)

1. Fill out information at the top of the sheets completely, including names of students involved in the testing. This includes filling out each box on each page of the habitat forms, so that if the sheets get separated, they can be individually identified.
2. Site codes for all sites can be found on the “Site Code List” handout.
3. At least 3 replicates are needed for all tests to be able to report the results in the database.
4. See separate handout on the statistical Q-test for determining if an outlying data point can be excluded from the average for a test.
5. For the nutrient tests, report the value as greater than the limit if the sample exceeds the limit on the test. The limit for each test is listed on the field instructions at the beginning of the procedure section.
6. Please make notes in Comments or open areas of the data sheet about any special circumstances in which samples were collected (e.g. from bridge using clean bucket) or if additional analyses were performed.

Turbidity:

1. Report turbidity as >60 cm if the symbol is already visible when the tube is completely full. The optional method using the DR850 colorimeter could be used to get a more quantitative value if this happens.
2. If second depth reading (screw is visible) is less than 60 cm, use 60 cm for first depth reading to compute the average for that observation rep.

General Comments for Nutrients:

1. All vials should be cleaned, acid washed and rinsed before storing for next analysis session. This will ensure that they are ready the next time you go to the field.
2. The preferred order of tests is phosphorus, ammonia, and then nitrate.
3. Based on measurements of standard solutions, the default calibration curve built into the DR850 colorimeters was adjusted July 2004 for the phosphorus and low range nitrate tests.
4. Please send us your students’ results from running standards that we give you. Run these one to two times a year with your students.
5. This summer we would like you to collect a second sample for us to run at the GBMSD lab. Let us know when you will be going out. Keep the sample on ice. We will need to arrange for shipping/transportation.
6. Use Q-test to remove outliers from your data.
7. If time permits, please rerun the sample if precision target is not met and there is not a legitimate reason to not include an observation. We will be revising our database to clearly separate data that meets QA/QC guidelines during the next year.